



# Effect of supplementary lighting on nutrient recovery of *Ocimum basilicum* and fishes in a polyculture aquaponic system

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## Abstract

The effects of prolonged photoperiod (additional night lighting) were investigated on the production of *Ocimum basilicum* (lemon basil) and fish/crayfish raised in a nutrient film technique aquaponic system. Hybrid lemon fin barb and red claw crayfish juveniles were co-cultured with *O. basilicum* and subjected to 12 h of ambient natural daylight and additional 0, 4, 8, and 12 h of night lighting for 14 weeks (two batches of 7 weeks each). The water quality parameters and the performance characteristics of the fish/crayfish/plant were evaluated, and the nutrient uptake (nitrogen, phosphorus, and potassium) in the system by the three organisms was also measured. The fish growth performance showed no significant differences ( $p > 0.05$ ) in all the growth parameters measured across the treatments. The body proximate and nitrogen, phosphate, and potassium (NPK) composition of the fish and crayfish were not significantly ( $p > 0.05$ ) different across the treatments. Similarly, there was no significant difference ( $p > 0.05$ ) in the NPK uptake by the fish and crayfish across the treatments. The plant growth performance showed that there were no significant ( $p > 0.05$ ) differences in the plant growth parameters measured, except for % plant height gained in the 12 h-light treatment ( $403.2 \pm 26.0\%$ ) which was significantly ( $p < 0.05$ ) higher than other treatments and cycles. Hence, this study demonstrated that varying supplementary night lighting has no significant effect on the growth performance of *O. basilicum*, hybrid lemon fin barb, and red claw crayfish. This study, therefore, suggests that supplementary night lighting should not be considered for *O. basilicum* production as it does not significantly improve the performance characteristics of the plant nor the fish co-cultured with it in a polyculture aquaponic system.

**Keywords:** Polyculture, Aquaponics, *Ocimum basilicum*, Nutrient recovery, Supplementary light

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## Introduction

Plants generally require an adequate amount of light, temperature, water, nutrients, and gases for optimal growth and development. Light is one of the most important abiotic factors and form of energy (radiant energy), can therefore regulate plant behaviour. Depending on the direction, quality, quantity, and duration of photoperiod, the development of plants can be greatly influenced (Dou et al., 2018). In many studies, the quality, and mass increase of plants have been linked to extended photoperiod regimes (Fernández-Cabanás et al., 2020; Liang & Chien, 2013, 2015). Photosynthesis and photomorphogenesis are examples of the primary and secondary metabolic responses that would be affected by the manipulation of photoperiod regimes (Ouzounis et al., 2015). Hence, photoperiod manipulation is key to producing consistent and predictable biomass in plant production systems (Fernández-Cabanás et al., 2020).

The quest for a more sustainable environment, efficient food production system, and food security are some of the reasons for the widespread interest in polyculture aquaponic systems (Hasan et al., 2017; Nuwansi et al., 2017). In contrast with conventional aquaponic systems which are mostly carried out with total manipulation of light under a controlled environment, the plants in a polyculture aquaponic system under a semi-controlled environment may require supplementary night lighting because of their saturated nutrients. Although a minimum of 8 to 10 h of lighting has been suggested for the optimal growth of plants in conventional systems (Malayeri et al., 2011), there seems to be a paucity of information on the response of different plants to different supplementary night lighting in a polyculture aquaponic system. However, the reason for the scarcity could be that aquaponics, or polyculture aquaponic systems, is a relatively recent technology that is only now starting to gain traction globally.

The main role of the plants in the aquaponic system is to significantly reduce the nutrient loading of the water as occasioned by the decomposing uneaten feeds and byproducts excreted by the fish (Love et al., 2015; Rakocy, 2012). The nutrients accumulated by plants in the system are mainly nitrogen and phosphorus, which are the principal compounds responsible for eutrophication in many aquatic environments (Liang & Chien, 2015; Petrucio & Esteves, 2000). Since the availability of light affects the photosynthetic activities of plants, it could be hypothesized that the manipulation of the photoperiod regime may also affect the nutrient removal capacity

of the plants connected to the system (Liang & Chien, 2013). *Ocimum basilicum* is a plant that can be used as an herb as well as an essential oil in cosmetics or pharmaceuticals (Ahmed et al., 2019). Thus, the current study is aimed at improving the performance of this important herb by increasing the lighting hours at night. This study is, therefore, designed to investigate the effects of supplementary night lighting on the growth and nutrient uptake by fish/crayfish and *O. basilicum* grown in a polyculture aquaponic system.

## Materials and Methods

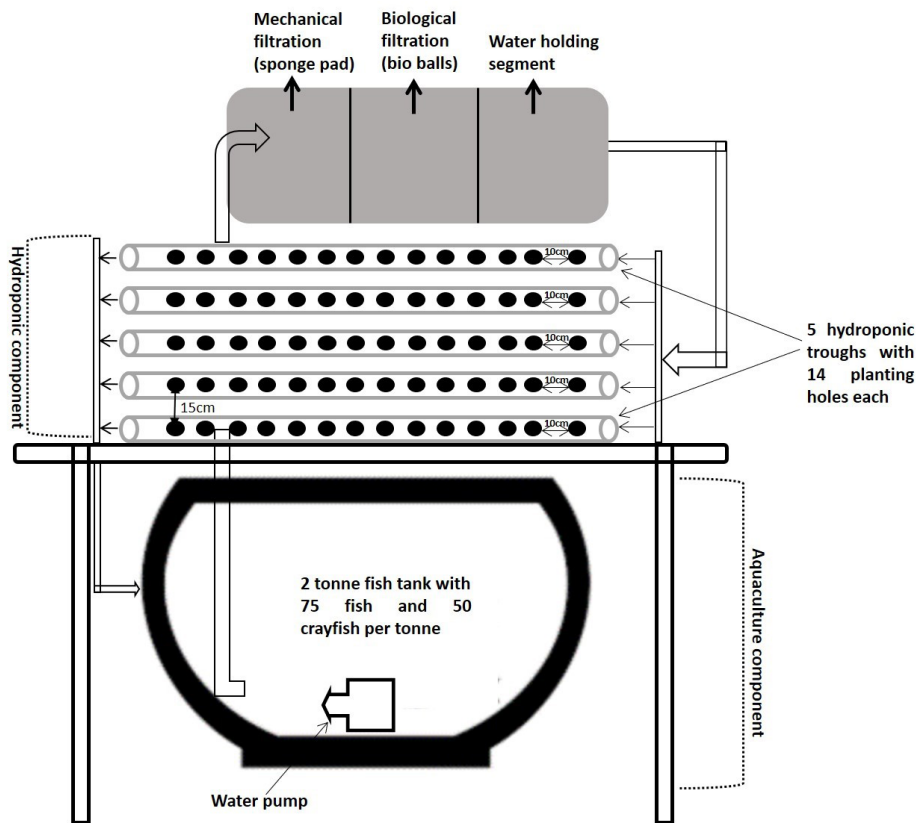
### Experimental setup, fish, crayfish, and *Ocimum basilicum*

The experiment was conducted at the Aquaculture Experimental Station, Universiti Putra Malaysia, Malaysia. The experimental setup consisted of 12-individual polyculture aquaponic units situated under a large open-shed aquaculture facility with natural light interception but enough for plant growth ( $57.74 \pm 1.43 \mu\text{mol m}^{-2}\text{s}^{-1}$ ). Each polyculture aquaponic unit consisted of a nutrient film technique vegetable trough unit made of 5 unplasticized polyvinyl chloride rectangular down-pipe rain gutters ( $5.95 \times 0.15 \times 0.07$  m), a filtration box, and a 2-tonne fibre glass fish tank. Each row held 14 pots, that is, 70 plant pots per culture unit. A portable (135-watt, 4,000 L/h) aquarium pump was used to pump water for recirculation in the system (Fig. 1). Each tank was filled with 1 tonne of dechlorinated state tap water.

The red claw crayfish was procured from Greeny Aquaculture, Johor, and the hybrid lemon fin barb fingerlings were from Jerantut Aquaculture Extension Centre in Perlok Pahang, Malaysia. The fish and crayfish were randomly stocked at 75 fish/50 crayfish/tonne and acclimatized to the polyculture aquaponic system for 4 weeks before *O. basilicum* seedlings were introduced into the system. Two 3-day-old seedlings of *O. basilicum* were (germinated on Petri dishes) transferred individually to each pot of the vegetable trough. During the study period of 14 weeks, the red claw crayfish and hybrid lemon fin barb fish were fed (5% body weight) commercial pellets (Blanca shrimp feeds, 36% crude protein, and Cargill starter, 34% crude protein, respectively).

### Supplementary lighting

Only the plants were subjected to supplementary lighting using the following four treatments: 12 h daylight and 0 h supplementary night light (control); 12 h daylight and 4 h supplementary night light; 12 h daylight and 8 h supplementary



**Fig. 1. Schematic representation of the aquaponic setup.**

night light; and 12 h natural daylight and 12 h supplementary night light. LED plant grow lights (Philips T8 tube, Philips Lighting, Bridgewater, NJ, USA) were used to provide artificial lighting in the range of  $650 \pm 50 \mu\text{mol m}^{-2}\text{s}^{-1}$ . Automatic timers were used to ensure strict adherence to the light regimes intended for each treatment. The plants were grown for 2 cycles, with each lasting for 7 weeks.

### Water quality analysis

Water quality parameters were taken every week from the effluent (water from the fish tank) and influent (water returning to the fish tank after going through the plant trough and filter box). Parameters such as dissolved oxygen (DO), temperature, electrical conductivity (EC), and pH were recorded *in situ* using a multi-parameter water probe (YSI 556 MPS, YSI, Yellow Springs, OH, USA). Thereafter, water samples were collected and used for the analysis of ammonia, nitrate, nitrite, phosphate, and potassium ions. The process involved filtration of the water samples in Whatman® filter paper (pore size 0.45  $\mu\text{m}$  and 47

mm diameter) before use. The ammonia and ammonium levels were then read using ammonia low range portable photometer (HI 96700, Hanna Instruments, Smithfield, RI, USA). However, the phosphate and potassium ion concentrations were analyzed using atomic absorption spectroscopy (TruMac Series, Leco, St. Joseph, MO, USA) as described by Ogah et al. (2020a).

### Evaluation of fish, crayfish, and plant performance

The fish and crayfish in this study were sampled weekly until the 14th week of culture (which was equivalent to the two cycles of the vegetables) to determine their performance. Hence, the fish and crayfish from each tank were individually batched and weighed to get the bulk weight and survival rate weekly. The information was also used to adjust the feed ration according to the new mean body weight obtained weekly. Thereafter, the weight gain, specific growth rate, average daily feed intake, feed conversion ratio (FCR), protein efficiency ratio (PER), and percentage survival were evaluated according to the equations described below.

$$\text{Growth rate (g/d)} = \frac{W_2 - W_1}{t_2 - t_1}$$

Where  $W_1$  = initial weight (g),  $W_2$  = final weight (g),  
 $t_2 - t_1$  = duration between  $W_2$  and  $W_1$  (days).

$$\text{Specific growth rate (\%/day)} = \frac{\log_e(W_2) - \log_e(W_1)}{t_2 - t_1} \times 100$$

$$\text{FCR} = \frac{\text{dryfeed consumed}}{W_2 - W_1}$$

$$\text{Feed conversion efficiency (\%FER)} = \frac{(W_2 - W_1) \times 100}{\text{dryfeed consumed}}$$

$$\text{PER} = \frac{\text{Fish weight gain (g)}}{\text{Total protein intake (g)}}$$

$$\text{Protein gained} = (\text{Weight}_{\text{final}} \times \% \text{Protein}_{\text{final}}) - (\text{Weight}_{\text{initial}} \times \% \text{Protein}_{\text{initial}})$$

$$\text{Lipid gained} = (\text{Weight}_{\text{final}} \times \% \text{Lipid}_{\text{final}}) - (\text{Weight}_{\text{initial}} \times \% \text{Lipid}_{\text{initial}})$$

$$\% \text{ Survival} = \frac{\text{fish stocked} - \text{mortality}}{\text{fish stocked}} \times 100$$

In this study, the evaluation of plant performance was based on the parameters such as leaf number (the number of true leaves in each pot), leaf area (area of 50 true leaves sampled from 25 plants) using a leaf area meter (LI-COR LI-3100C, Lincoln, NE, USA), plant height (i.e., using a meter rule), and chlorophyll content (Konica Minolta SPAD 502 Plus, Tokyo, Japan). These parameters were measured at the beginning (during transplanting) and at the end of the experiment.

### Fish, crayfish, and plant tissue analysis

The proximate composition of the fish and crayfish in the polyculture aquaponics system was done according to the method specified by AOAC (1997). Also, the plant leaf and the fish/crayfish tissues were analyzed for nitrogen (N), phosphate (P), and potassium (K) to estimate the initial and final values before and after the study. This was done using the Perkin Elmer Analyst 400 spectrophotometer flame method (Bader, 2011). The nutrient retention in the plants was evaluated using the following

equations:

$$\text{Nutrient retention} = 100 \times \frac{\text{Nutrient gained in plant or fish}}{\text{Total nutrient in feed given}}$$

$$\text{Nutrient uptake} = (\text{Final weight} \times \text{Final NPK content}) - (\text{Initial weight} \times \text{Initial NPK content})$$

### Statistical analysis

Descriptive statistics of the collected data were done using Minitab 14 to obtain the means and standard errors for the different data sets in this study. The data were tested for normality and homogeneity of variance before they were subjected to an analysis of variance (ANOVA). In particular, the water quality parameter data were analyzed using a 2-way ANOVA and the results were presented in graphs. The fish, crayfish, and plant growth performances and body/nutrient compositions were analyzed using one-way ANOVA. Also, it should be noted that the percentage data for nutrient recovery were arcsine transformed.

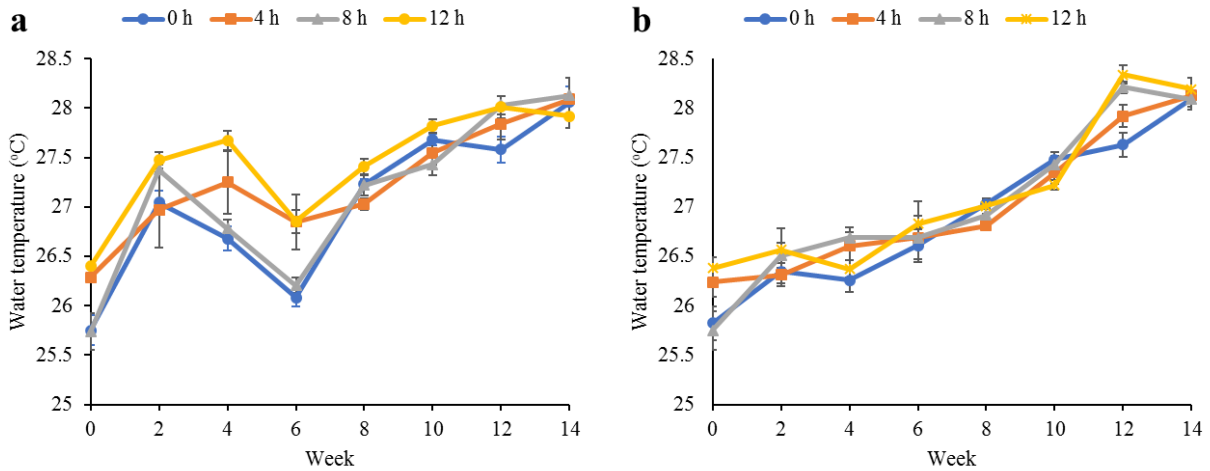
## Results

### Water quality

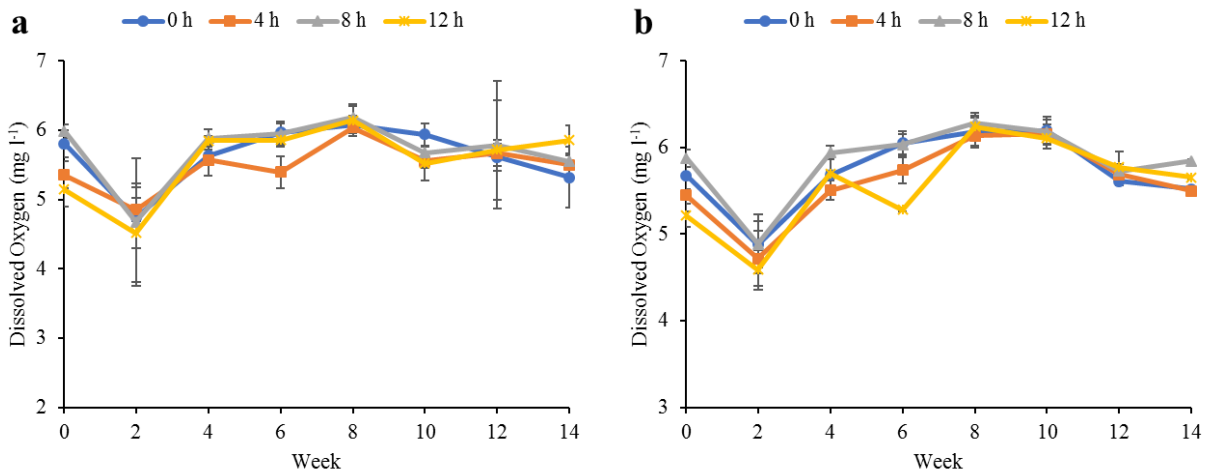
Water quality parameters in the culture tanks and plant trough outlets throughout the experiment are shown in Figs. 2–6. Generally, there were no significant differences ( $p > 0.05$ ) in the water parameters observed between the culture tanks and plant troughs. The water temperature in the culture tanks and trough increased steadily while it ranged from 25.7°C–28.1°C. However, the DO, pH, and EC ranges obtained in this study were 4.5–6.3 mg/L, 5.8–7.2 and 0.2–0.75 mS/cm respectively. In both culture tanks and plant troughs, the ammonia-nitrogen was within the safe limit (0.2–1.4 mg/L) for the culture of fish and crayfish, while there were no significant differences ( $p > 0.05$ ) in the ammonia concentration across the different treatments.

### Fish/crayfish growth performance and body composition

The growth indices and survival of hybrid lemon fin barb and red claw crayfish cultured with *O. basilicum* under different night lighting regimes are shown in Table 1. The fish growth performance showed that there were no significant differences ( $p > 0.05$ ) in all the growth parameters assessed across the treatments. In addition, the growth performance of the crayfish showed 20.17–22.57 g weight gain, 2.34–2.53 FCR, and 1.17–



**Fig. 2.** Water temperature in (A) culture tanks and (B) plant trough outlet where hybrid lemon fin barb and red claw crayfish were cultured with basil and jute mallow herbs exposed to supplementary night lighting in a polyculture aquaponic system.



**Fig. 3.** Dissolved oxygen in (A) culture tanks and (B) plant trough outlet where hybrid lemon fin barb and red claw crayfish were cultured with basil and jute mallow herbs exposed to supplementary night lighting in a polyculture aquaponic system.

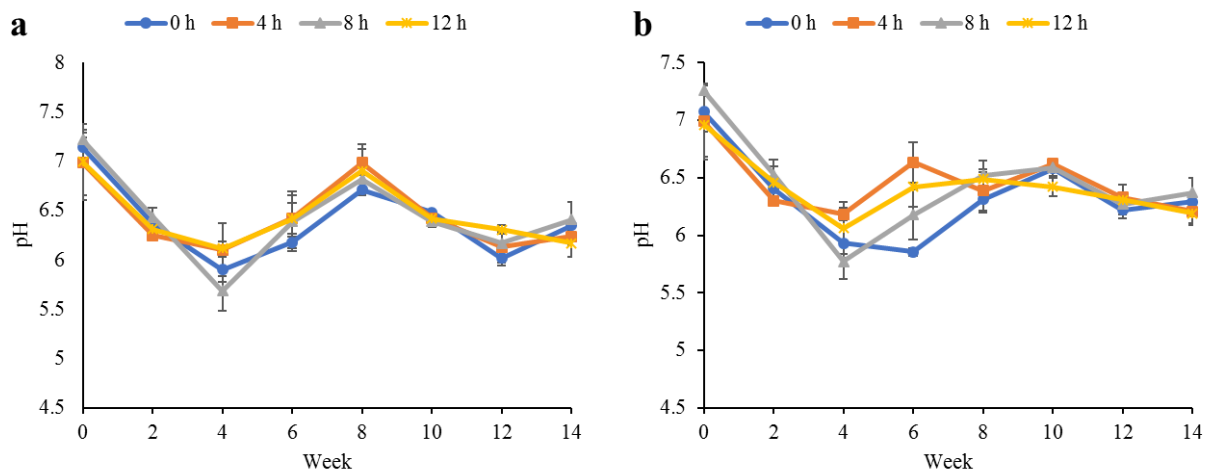
1.26 PER. However, no significant differences ( $p > 0.05$ ) were observed in growth parameters measured across the treatments in either fish or crayfish.

The body proximate composition of the fish and crayfish was not significantly ( $p > 0.05$ ) different across the treatments (Table 2). However, the NPK composition of the fish and crayfish was not significantly ( $p > 0.05$ ) different across the treatments (Table 3). In the fish carcass, NPK values obtained were 2.93–2.97 mg/g, 0.56–0.62 mg/g, and 0.26–0.28 mg/g, respectively, whereas

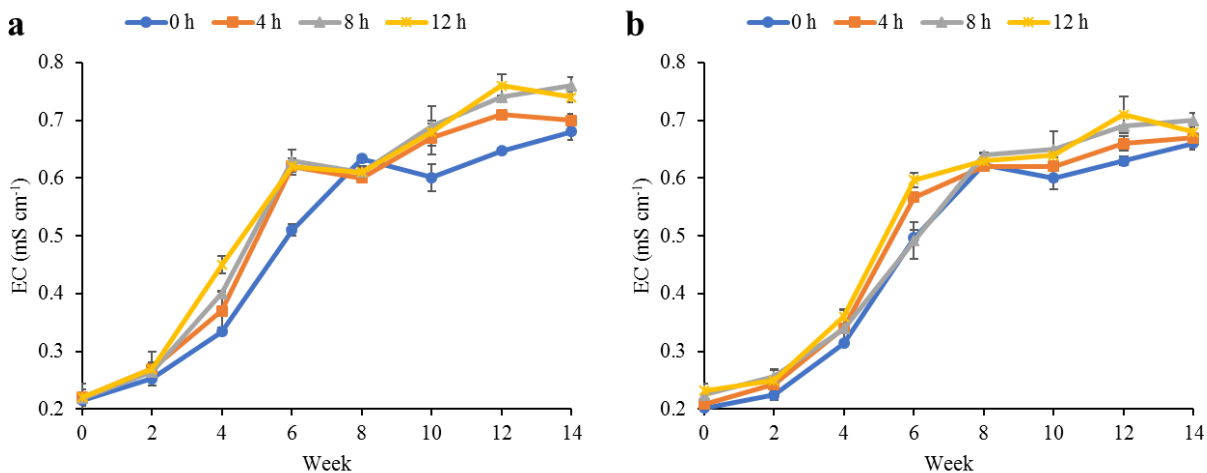
N values of 2.60–2.65 mg/g, P values of 0.15–0.16 mg/g, and K values of 0.05–0.06 mg/g were obtained in the carcass of crayfish.

**Growth performance and composition of *Ocimum basilicum***

Table 4 shows the summary of plant performances in different extended night lighting regimes during two culture cycles. Overall, there were no significant ( $p > 0.05$ ) differences in the plant growth parameters measured, except for % plant height gained in the 12 h-light treatment, which was significantly ( $p <$



**Fig. 4.** pH in (A) culture tanks and (B) plant trough outlet where hybrid lemon fin barb and red claw crayfish were cultured with basil and jute mallow herbs exposed to supplementary night lighting in a polyculture aquaponic system.



**Fig. 5.** Electrical conductivity in (A) culture tanks and (B) plant trough outlet where hybrid lemon fin barb and red claw crayfish were cultured with basil and jute mallow herbs exposed to supplementary night lighting in a polyculture aquaponic system.

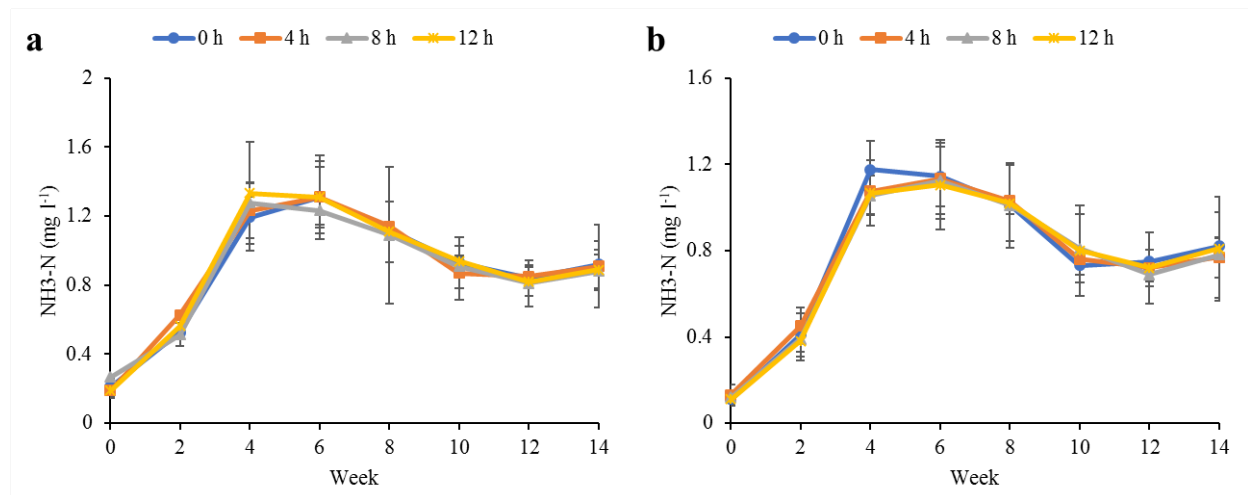
0.05) higher than other treatments and cycles.

Furthermore, *O. basilicum* nutrient compositions before and after being subjected to different lighting regimes are shown in Table 5. Similar trends in the composition of plants were observed in both plant cycles. There were no significant ( $p > 0.05$ ) differences in the final NPK of *O. basilicum* in both cycles.

**Nutrient uptake and recovery**

The nutrient uptake by the fish showed that N uptake ranges

from 250.0–259.1 g/culture unit, P from 46.4–51.4 g/culture unit, and K from 22.0–22.6 g/culture unit. There was no significant difference ( $p > 0.05$ ) in the NPK uptake by the fish across the treatments (Table 6). Similarly, there was no significant difference ( $p > 0.05$ ) in the NPK uptake by the crayfish across the treatments. The nutrient uptake in relation to the total system showed that there were no significant differences ( $p > 0.05$ ) across the treatments in the NPK uptake by *O. basilicum* and the total system.



**Fig. 6. Ammonia-nitrogen in (A) culture tanks and (B) plant trough outlet where hybrid lemon fin barb and red claw crayfish were cultured with basil and jute mallow herbs exposed to supplementary night lighting in a polyculture aquaponic system.**

**Table 1. Growth performance of hybrid lemon fin barb and red claw crayfish in a polyculture aquaponic system with *Ocimum basilicum* exposed to different light regimen for 14 weeks**

Performance	Fish				Crayfish			
	0 h	4 h	8 h	12 h	0 h	4 h	8 h	12 h
IBW (g)	12.3 ± 0.61	12.4 ± 0.17	12.2 ± 0.74	12.57 ± 1.13	4.1 ± 0.32	4.1 ± 0.15	4.23 ± 0.09	4.13 ± 0.32
FBW (g)	124.87 ± 4.39 <sup>a</sup>	113.87 ± 7.91 <sup>a</sup>	128.27 ± 7.24 <sup>a</sup>	127.93 ± 3.68 <sup>a</sup>	24.27 ± 3.26 <sup>x</sup>	25.11 ± 3.26 <sup>x</sup>	26.53 ± 3.14 <sup>x</sup>	26.7 ± 1.79 <sup>x</sup>
MWG (g)	112.57 ± 4.61 <sup>a</sup>	113.87 ± 7.83 <sup>a</sup>	116.07 ± 7.74 <sup>a</sup>	115.37 ± 4.75 <sup>a</sup>	20.17 ± 2.96 <sup>x</sup>	21.01 ± 3.12 <sup>x</sup>	22.3 ± 3.09 <sup>x</sup>	22.57 ± 1.47 <sup>x</sup>
SGR (%/d)	2.21 ± 0.07 <sup>a</sup>	2.21 ± 0.05 <sup>a</sup>	2.24 ± 0.1 <sup>a</sup>	2.21 ± 0.12 <sup>a</sup>	1.68 ± 0.06 <sup>x</sup>	1.71 ± 0.1 <sup>x</sup>	1.73 ± 0.11 <sup>x</sup>	1.78 ± 0.01 <sup>x</sup>
DFI (%/d)	3.05 ± 0.13 <sup>a</sup>	2.99 ± 0.05 <sup>a</sup>	3.02 ± 0.06 <sup>a</sup>	3.07 ± 0.13 <sup>a</sup>	3.64 ± 0.09 <sup>x</sup>	3.56 ± 0.12 <sup>x</sup>	3.52 ± 0.09 <sup>x</sup>	3.50 ± 0.14 <sup>x</sup>
FCR	1.82 ± 0.09 <sup>a</sup>	1.79 ± 0.04 <sup>a</sup>	1.80 ± 0.07 <sup>a</sup>	1.84 ± 0.05 <sup>a</sup>	2.53 ± 0.05 <sup>x</sup>	2.45 ± 0.13 <sup>x</sup>	2.42 ± 0.21 <sup>x</sup>	2.34 ± 0.09 <sup>x</sup>
PER	1.62 ± 0.08 <sup>a</sup>	1.65 ± 0.04 <sup>a</sup>	1.64 ± 0.06 <sup>a</sup>	1.61 ± 0.04 <sup>a</sup>	1.17 ± 0.02 <sup>x</sup>	1.21 ± 0.06 <sup>x</sup>	1.23 ± 0.10 <sup>x</sup>	1.26 ± 0.05 <sup>x</sup>
Survival (%)	97.33 ± 0.77 <sup>a</sup>	97.78 ± 0.45 <sup>a</sup>	96.44 ± 0.9 <sup>a</sup>	95.56 ± 2.47 <sup>a</sup>	84.67 ± 4.81 <sup>x</sup>	84.67 ± 2.4 <sup>x</sup>	86.67 ± 0.67 <sup>x</sup>	85.33 ± 0.67 <sup>x</sup>
Total P (kg/tonne)	9.11 ± 0.25 <sup>a</sup>	9.26 ± 0.55 <sup>a</sup>	9.28 ± 0.58 <sup>a</sup>	9.18 ± 1.13 <sup>a</sup>	1.04 ± 0.20 <sup>x</sup>	1.07 ± 0.17 <sup>x</sup>	1.15 ± 0.13 <sup>x</sup>	1.14 ± 0.08 <sup>x</sup>

Data are represented as mean ± SE.

<sup>a,x</sup> Values within the same row and species with different superscript letters are significantly different ( $p < 0.05$ ).

IBW, initial body weight; FBW, final body weight; MWG, mean weight gain; SGR, specific growth rate; DFI, daily feed intake; FCR, feed conversion ratio; PER, protein efficiency ratio; Total P: total production.

The percentage nutrient recovery of hybrid lemon fin barb and red claw crayfish cultured with *O. basilicum* under different night lighting regimes is shown in Table 7. There were no significant differences ( $p > 0.05$ ) across the treatments in NPK recovery from both fish and crayfish. However, the nutrient recovered from *O. basilicum* in both cycles was similar and there were no significant differences ( $p > 0.05$ ) in % NPK recovered across the treatments in both cycles. The system total P recovered from 4–12 h-treatments was significantly higher ( $p < 0.05$ ) than the system total P recovered.

## Discussion

Water quality parameters are very important variables of interest in a polyculture aquaponic system for optimum growth of fish and plants (Martan, 2008; Oladimeji et al., 2020). This is because water quality imbalances such as high ammonia levels could be detrimental to fishes raised in such a closed aquaculture system. The levels of most water qualities observed in the current study are in line with the reports of Boyd & Gross (2000) and Yildiz et al. (2017) and are considered safe for the culture of experimental

**Table 2. Body composition of hybrid lemon fin barb and red claw crayfish (% wet weight) grown with *Ocimum basilicum* exposed to different night lighting**

Treatments	Nutrients (%)					
	Moisture	Crude protein	Crude lipid	Ash	Crude fibre	NFE
<b>Fish</b>						
Initial	75.36 ± 0.56	12.6 ± 0.29	4.53 ± 0.38	2.05 ± 0.03	0.10 ± 0.0	5.38 ± 1.16
0 h	67.9 ± 0.22 <sup>a</sup>	18.41 ± 0.26 <sup>a</sup>	4.10 ± 0.07 <sup>a</sup>	2.25 ± 0.09 <sup>a</sup>	0.16 ± 0.02 <sup>a</sup>	7.19 ± 0.06 <sup>a</sup>
4 h	67.84 ± 0.84 <sup>a</sup>	18.33 ± 0.41 <sup>a</sup>	4.18 ± 0.22 <sup>a</sup>	2.27 ± 0.11 <sup>a</sup>	0.17 ± 0.02 <sup>a</sup>	7.18 ± 0.56 <sup>a</sup>
8 h	67.59 ± 0.66 <sup>a</sup>	18.58 ± 0.50 <sup>a</sup>	4.05 ± 0.05 <sup>a</sup>	2.36 ± 0.30 <sup>a</sup>	0.21 ± 0.02 <sup>a</sup>	7.21 ± 0.34 <sup>a</sup>
12 h	67.79 ± 0.69 <sup>a</sup>	18.55 ± 0.32 <sup>a</sup>	4.1 ± 0.06 <sup>a</sup>	2.35 ± 0.23 <sup>a</sup>	0.21 ± 0.03 <sup>a</sup>	7.01 ± 0.52 <sup>a</sup>
<b>Crayfish</b>						
Initial	78.88 ± 0.23	12.63 ± 0.28	1.04 ± 0.01	1.02 ± 0.05	0.26 ± 0.02	6.17 ± 0.10
0 h	75.33 ± 0.60 <sup>x</sup>	16.23 ± 0.28 <sup>x</sup>	1.06 ± 0.02 <sup>x</sup>	1.13 ± 0.01 <sup>x</sup>	0.12 ± 0.01 <sup>x</sup>	6.12 ± 0.33 <sup>x</sup>
4 h	75.34 ± 0.25 <sup>x</sup>	16.29 ± 0.28 <sup>x</sup>	1.07 ± 0.02 <sup>x</sup>	1.2 ± 0.06 <sup>x</sup>	0.11 ± 0.01 <sup>x</sup>	5.99 ± 0.18 <sup>x</sup>
8 h	74.95 ± 0.44 <sup>x</sup>	16.54 ± 0.44 <sup>x</sup>	1.05 ± 0.04 <sup>x</sup>	1.07 ± 0.02 <sup>x</sup>	0.11 ± 0.06 <sup>x</sup>	6.25 ± 0.78 <sup>x</sup>
12 h	75.02 ± 0.46 <sup>x</sup>	16.52 ± 0.44 <sup>x</sup>	1.03 ± 0.07 <sup>x</sup>	1.08 ± 0.03 <sup>x</sup>	0.12 ± 0.01 <sup>x</sup>	6.23 ± 0.79 <sup>x</sup>

Data are represented as mean ± SE.

<sup>a,x</sup> Values within the same column and species with different superscript letters are significantly different ( $p < 0.05$ ).

**Table 3. Fish and crayfish NPK composition in a polyculture aquaponic system exposed to different light regimen (wet weight basis)**

Parameter	Fish				Crayfish			
	0 h	4 h	8 h	12 h	0 h	4 h	8 h	12 h
Initial N (mg/g)	2.06 ± 0.05	2.02 ± 0.05	2.02 ± 0.05	2.03 ± 0.04	2.02 ± 0.01	2.02 ± 0.02	2.03 ± 0.01	2.04 ± 0.02
Final N (mg/g)	2.95 ± 0.04 <sup>a</sup>	2.93 ± 0.07 <sup>a</sup>	2.97 ± 0.08 <sup>a</sup>	2.97 ± 0.05 <sup>a</sup>	2.60 ± 0.05 <sup>x</sup>	2.61 ± 0.03 <sup>x</sup>	2.65 ± 0.07 <sup>x</sup>	2.64 ± 0.07 <sup>x</sup>
Initial P (mg/g)	0.53 ± 0.02	0.57 ± 0.01	0.58 ± 0.0	0.59 ± 0.01	0.11 ± 0.02	0.10 ± 0.02	0.11 ± 0.01	0.11 ± 0.02
Final P (mg/g)	0.56 ± 0.04 <sup>a</sup>	0.60 ± 0.01 <sup>a</sup>	0.60 ± 0.01 <sup>a</sup>	0.62 ± 0.01 <sup>a</sup>	0.16 ± 0.01 <sup>x</sup>	0.16 ± 0.0 <sup>x</sup>	0.15 ± 0.0 <sup>x</sup>	0.16 ± 0.01 <sup>x</sup>
Initial K (mg/g)	0.25 ± 0.0	0.25 ± 0.01	0.25 ± 0.02	0.26 ± 0.01	0.04 ± 0.01	0.04 ± 0.02	0.04 ± 0.0	0.04 ± 0.01
Final K (mg/g)	0.27 ± 0.01 <sup>a</sup>	0.26 ± 0.01 <sup>a</sup>	0.27 ± 0.01 <sup>a</sup>	0.28 ± 0.0 <sup>a</sup>	0.06 ± 0.0 <sup>x</sup>	0.06 ± 0.01 <sup>x</sup>	0.06 ± 0.0 <sup>x</sup>	0.05 ± 0.01 <sup>x</sup>

Data are represented as mean ± SE.

<sup>a,x</sup> Values within the same row and species with different superscript letters are significantly different ( $p < 0.05$ ).

N, nitrogen; P, phosphate; K, potassium.

organisms (crayfish and fish) used. Generally, the photoperiods have no significant effect on the water qualities observed in this study across the treatments, and this is similar to the reports by Liang & Chien (2013) and Ogah et al. (2020b). The decrease in pH levels in the culture tanks and plant trough outlets under the different light regimes could be attributed to nitrate accumulation in the biofilter during the culture period (Boyd & Gross, 2000; Oommen et al., 2019). Also, the levels and variations of the EC observed in this study were higher than in the earlier study by Ogah et al. (2020b) which reported 0.25–0.42 mS/cm in a similar polyculture aquaponic system. Maintaining high DO in a bicultural aquaponic system cannot be over-emphasized as

it helps with the breakdown of organic materials in the culture water, therefore making available sufficient nutrients for plant absorption and improved health (Eck et al., 2019; Rakocy et al., 2006). The current study also observed that the DO for the polyculture aquaponic system used was within the recommended range for aquaculture purposes (Yildiz et al., 2017) and may have been made possible because of the provision of aeration mechanisms that supply additional DO to the system. The water temperature observed in this study falls within the optimum range for fish growth in aquaponics (Yildiz et al., 2017) and is also similar to the report by Ogah et al. (2020b) which reported 27°C–29°C in a similar aquaponic system. However, there was a



**Table 4. *Ocimum basilicum* characteristics exposed to 12 h daylight, followed by different lengths of night lighting**

Parameter	Cycle 1				Cycle 2			
	0 h	4 h	8 h	12 h	0 h	4 h	8 h	12 h
Initial LA (cm <sup>2</sup> /leaf)	2.4 ± 0.2	2.4 ± 0.1	2.4 ± 0.2	2.4 ± 0.2	2.4 ± 0.0	2.4 ± 0.0	2.4 ± 0.2	2.4 ± 0.2
Final LA (cm <sup>2</sup> /leaf)	15.5 ± 0.3 <sup>a</sup>	15.7 ± 0.7 <sup>a</sup>	15.7 ± 0.3 <sup>a</sup>	15.8 ± 0.7 <sup>a</sup>	16.2 ± 0.5 <sup>a</sup>	16.3 ± 0.5 <sup>a</sup>	16.3 ± 0.8 <sup>a</sup>	16.2 ± 0.8 <sup>a</sup>
LA gain (cm <sup>2</sup> /leaf)	13.1 ± 0.2 <sup>a</sup>	13.3 ± 0.7 <sup>a</sup>	13.3 ± 0.1 <sup>a</sup>	13.4 ± 0.5 <sup>a</sup>	13.8 ± 0.4 <sup>a</sup>	13.9 ± 0.5 <sup>a</sup>	13.9 ± 0.6 <sup>a</sup>	13.9 ± 0.7 <sup>a</sup>
LA gain (%/leaf)	549.8 ± 31.9 <sup>a</sup>	550.7 ± 13.7 <sup>a</sup>	526.9 ± 42.7 <sup>a</sup>	555.6 ± 25.0 <sup>a</sup>	587.1 ± 10.9 <sup>a</sup>	588.5 ± 11.0 <sup>a</sup>	589.5 ± 20.6 <sup>a</sup>	586.3 ± 13.8 <sup>a</sup>
Initial LN (leaves/pot)	6.0 ± 0.6	6.3 ± 0.7	6.0 ± 1.2	6.7 ± 0.7	6.7 ± 0.3	6.3 ± 0.7	6.7 ± 1.3	7.0 ± 0.6
Final LN (leaves/pot)	28.7 ± 0.9 <sup>a</sup>	31.0 ± 3.1 <sup>a</sup>	30.3 ± 4.4 <sup>a</sup>	31.3 ± 2.8 <sup>a</sup>	33.3 ± 2.4 <sup>a</sup>	33.7 ± 3.0 <sup>a</sup>	34.3 ± 4.7 <sup>a</sup>	35.3 ± 3.4 <sup>a</sup>
LN gain (leaves/pot)	22.7 ± 0.9 <sup>a</sup>	24.7 ± 2.4 <sup>a</sup>	24.3 ± 3.3 <sup>a</sup>	24.7 ± 2.2 <sup>a</sup>	26.7 ± 2.2 <sup>a</sup>	27.3 ± 2.3 <sup>a</sup>	27.7 ± 3.3 <sup>a</sup>	28.3 ± 2.8 <sup>a</sup>
LN gain (%)	386.2 ± 47.3 <sup>a</sup>	420.8 ± 53.2 <sup>a</sup>	420.8 ± 53.2 <sup>a</sup>	370.8 ± 6.4 <sup>a</sup>	400.0 ± 24.7 <sup>a</sup>	434.3 ± 17.8 <sup>a</sup>	433.3 ± 45.8 <sup>a</sup>	404.4 ± 16.6 <sup>a</sup>
Initial PH (cm/pot)	5.1 ± 0.4	5.7 ± 0.1	5.4 ± 0.4	5.5 ± 0.2	5.3 ± 0.4	5.3 ± 0.2	5.3 ± 0.3	5.4 ± 0.1
Final PH (cm/pot)	21.8 ± 1.4 <sup>a</sup>	24.9 ± 1.6 <sup>a</sup>	26.3 ± 2.1 <sup>a</sup>	26.1 ± 1.8 <sup>a</sup>	25.6 ± 1.7 <sup>a</sup>	26.2 ± 2.2 <sup>a</sup>	26.6 ± 1.8 <sup>a</sup>	26.1 ± 1.8 <sup>a</sup>
PH gain (cm/pot)	16.6 ± 1.4 <sup>a</sup>	19.2 ± 1.6 <sup>a</sup>	20.8 ± 1.8 <sup>a</sup>	21.7 ± 0.8 <sup>a</sup>	20.3 ± 1.3 <sup>a</sup>	20.9 ± 2.0 <sup>a</sup>	21.3 ± 1.6 <sup>a</sup>	21.8 ± 0.8 <sup>a</sup>
PH gain (%)	329.3 ± 40.9 <sup>b</sup>	338.7 ± 26.0 <sup>ab</sup>	382.7 ± 17.4 <sup>ab</sup>	394.4 ± 27.4 <sup>a</sup>	382.1 ± 13.1 <sup>ab</sup>	393.3 ± 29.0 <sup>a</sup>	397.9 ± 12.3 <sup>a</sup>	403.2 ± 26.0 <sup>a</sup>
Initial CC (mg cm <sup>-2</sup> )	9.3 ± 0.9	9.5 ± 0.1	9.5 ± 0.7	9.8 ± 0.7	9.8 ± 0.2	9.9 ± 0.3	9.8 ± 0.4	9.9 ± 0.3
Final CC (mg cm <sup>-2</sup> )	19.8 ± 1.4 <sup>a</sup>	20.2 ± 0.4 <sup>a</sup>	20.8 ± 1.3 <sup>a</sup>	20.6 ± 1.5 <sup>a</sup>	25.5 ± 1.8 <sup>a</sup>	25.2 ± 1.5 <sup>a</sup>	26.1 ± 2.9 <sup>a</sup>	25.6 ± 2.5 <sup>a</sup>
CC gain (mg cm <sup>-2</sup> )	10.5 ± 0.4 <sup>a</sup>	10.7 ± 0.3 <sup>a</sup>	11.3 ± 0.7 <sup>a</sup>	11.8 ± 1.2 <sup>a</sup>	15.6 ± 1.6 <sup>a</sup>	15.3 ± 1.2 <sup>a</sup>	16.3 ± 2.6 <sup>a</sup>	17.3 ± 1.1 <sup>a</sup>
CC gain (%)	115.4 ± 8.1 <sup>a</sup>	112.2 ± 2.5 <sup>a</sup>	119.8 ± 4.6 <sup>a</sup>	123.9 ± 22.0 <sup>a</sup>	158.5 ± 13.2 <sup>a</sup>	154.5 ± 8.6	164.8 ± 21.1 <sup>a</sup>	175.0 ± 15.8 <sup>a</sup>
Initial FW (g/unit culture)	70.5 ± 3.4	74.4 ± 5.2	71.2 ± 2.0	72.4 ± 3.5	73.1 ± 1.6	73.0 ± 4.7	73.2 ± 1.9	73.8 ± 3.8
Final FW (g/unit culture)	429.0 ± 17.1 <sup>a</sup>	444.2 ± 23.3 <sup>a</sup>	444.3 ± 16.6 <sup>a</sup>	442.2 ± 15.4 <sup>a</sup>	464.3 ± 5.4 <sup>a</sup>	465.5 ± 26.1 <sup>a</sup>	468.9 ± 19.1 <sup>a</sup>	466.5 ± 24.9 <sup>a</sup>
FW gain (g/unit culture)	358.5 ± 13.8 <sup>a</sup>	369.8 ± 19.7 <sup>a</sup>	373.1 ± 15.1 <sup>a</sup>	369.7 ± 12.1 <sup>a</sup>	391.2 ± 4.1 <sup>a</sup>	392.5 ± 21.5 <sup>a</sup>	395.7 ± 17.2 <sup>a</sup>	392.7 ± 21.1 <sup>a</sup>
FW gain (%)	509.2 ± 5.5 <sup>a</sup>	500.2 ± 28.6 <sup>a</sup>	524.0 ± 15.9 <sup>a</sup>	511.3 ± 10.4 <sup>a</sup>	535.2 ± 7.8 <sup>a</sup>	538.3 ± 9.0 <sup>a</sup>	540.1 ± 10.2 <sup>a</sup>	532.2 ± 1.6 <sup>a</sup>
Yield (g m <sup>-2</sup> )	286.0 ± 11.4 <sup>a</sup>	296.1 ± 15.1 <sup>a</sup>	296.2 ± 11.0 <sup>a</sup>	294.8 ± 10.3 <sup>a</sup>	309.5 ± 3.6 <sup>a</sup>	310.4 ± 17.4 <sup>a</sup>	312.6 ± 12.7 <sup>a</sup>	311.0 ± 16.6 <sup>a</sup>

<sup>ab</sup> Mean ± SE within the same row and growth cycle with different superscripts are significantly different at *p* < 0.05. LA, leaf area; LN, leaf number; PH, plant height; CC, chlorophyll content; FW, fresh weight.

**Table 5. NPK composition of *Ocimum basilicum* exposed to different night lighting regimen in a polyculture aquaponic system (wet weight basis)**

Parameter	0 h	4 h	8 h	12 h
Cycle 1				
Initial N (mg/g)	20.38 ± 0.24	20.41 ± 0.11	20.40 ± 0.13	20.40 ± 0.21
Final N (mg/g)	31.16 ± 0.15 <sup>a</sup>	31.07 ± 0.32 <sup>a</sup>	31.13 ± 0.03 <sup>a</sup>	31.15 ± 0.48 <sup>a</sup>
Initial P (mg/g)	0.57 ± 0.01	0.58 ± 0.02	0.59 ± 0.02	0.58 ± 0.02
Final P (mg/g)	1.12 ± 0.02 <sup>a</sup>	1.13 ± 0.02 <sup>a</sup>	1.13 ± 0.03 <sup>a</sup>	1.13 ± 0.03 <sup>a</sup>
Initial K (mg/g)	1.10 ± 0.01	1.09 ± 0.01	1.10 ± 0.02	1.10 ± 0.03
Final K (mg/g)	1.50 ± 0.01 <sup>a</sup>	1.49 ± 0.02 <sup>a</sup>	1.49 ± 0.03 <sup>a</sup>	1.50 ± 0.06 <sup>a</sup>
Cycle 2				
Initial N (mg/g)	20.85 ± 0.14	20.86 ± 0.18	20.83 ± 0.19	20.85 ± 0.14
Final N (mg/g)	32.22 ± 0.25 <sup>a</sup>	32.22 ± 0.11 <sup>a</sup>	32.11 ± 0.25 <sup>a</sup>	32.25 ± 0.06 <sup>a</sup>
Initial P (mg/g)	0.59 ± 0.02	0.58 ± 0.03	0.59 ± 0.02	0.59 ± 0.02
Final P (mg/g)	1.21 ± 0.01 <sup>a</sup>	1.21 ± 0.0 <sup>a</sup>	1.21 ± 0.01 <sup>a</sup>	1.23 ± 0.03 <sup>a</sup>
Initial K (mg/g)	1.09 ± 0.02	1.09 ± 0.03	1.09 ± 0.04	1.10 ± 0.03
Final K (mg/g)	1.51 ± 0.06 <sup>a</sup>	1.51 ± 0.08 <sup>a</sup>	1.53 ± 0.07 <sup>a</sup>	1.53 ± 0.01 <sup>a</sup>

Data are represented as mean ± SE.

<sup>a</sup>Values within the same row with different superscript letters are significantly different (*p* < 0.05). N, nitrogen; P, phosphate; K, potassium.

**Table 6. NPK total uptake (g/culture unit) in hybrid lemon fin barb, red claw crayfish, and *Ocimum basilicum* exposed to different night lighting regimen in a polyculture aquaponic system**

Nutrient uptake (g/culture unit)	0 h	4 h	8 h	12 h
Fish				
N	250.34 ± 10.34 <sup>a</sup>	253.36 ± 18.98 <sup>a</sup>	259.09 ± 25.03 <sup>a</sup>	254.14 ± 13.63 <sup>a</sup>
P	46.36 ± 0.65 <sup>a</sup>	50.21 ± 2.76 <sup>a</sup>	50.82 ± 3.73 <sup>a</sup>	51.38 ± 4.20 <sup>a</sup>
K	22.49 ± 0.89 <sup>a</sup>	22.02 ± 0.67 <sup>a</sup>	22.64 ± 0.21 <sup>a</sup>	22.45 <sup>s</sup> ± 0.90 <sup>a</sup>
Crayfish				
N	23.36 ± 4.27 <sup>a</sup>	24.44 ± 4.27 <sup>a</sup>	26.68 ± 3.55 <sup>a</sup>	26.59 ± 2.30 <sup>a</sup>
P	1.45 ± 0.36 <sup>a</sup>	1.47 ± 0.21 <sup>a</sup>	1.56 ± 0.16 <sup>a</sup>	1.61 ± 0.07 <sup>a</sup>
K	0.53 ± 0.11 <sup>a</sup>	0.55 ± 0.12 <sup>a</sup>	0.59 ± 0.1 <sup>a</sup>	0.60 ± 0.11 <sup>a</sup>
Total (fish and crayfish)				
N	273.71 ± 12.06 <sup>a</sup>	277.8 ± 17.78 <sup>a</sup>	285.76 ± 22.56 <sup>a</sup>	280.73 ± 11.73 <sup>a</sup>
P	47.81 ± 0.80 <sup>a</sup>	51.68 ± 2.65 <sup>a</sup>	52.38 ± 3.57 <sup>a</sup>	52.99 ± 4.13 <sup>a</sup>
K	23.01 ± 1.45 <sup>a</sup>	22.57 ± 0.73 <sup>a</sup>	23.24 ± 0.9 <sup>a</sup>	23.05 ± 1.15 <sup>a</sup>
<i>O. basilicum</i> (cycle 1)				
N	119.26 ± 4.07 <sup>a</sup>	122.86 ± 7.15 <sup>a</sup>	123.76 ± 4.51 <sup>a</sup>	122.77 ± 1.97 <sup>a</sup>
P	4.38 ± 0.16 <sup>a</sup>	4.56 ± 0.18 <sup>a</sup>	4.60 ± 0.07 <sup>a</sup>	4.58 ± 0.25 <sup>a</sup>
K	5.64 ± 0.21 <sup>a</sup>	5.79 ± 0.31 <sup>a</sup>	5.81 ± 0.11 <sup>a</sup>	5.83 ± 0.24 <sup>a</sup>
<i>O. basilicum</i> (cycle 2)				
N	134.36 ± 1.96 <sup>a</sup>	134.75 ± 7.47 <sup>a</sup>	135.31 ± 5.54 <sup>a</sup>	135.07 ± 7.14 <sup>a</sup>
P	5.20 ± 0.11 <sup>a</sup>	5.20 ± 0.28 <sup>a</sup>	5.24 ± 0.21 <sup>a</sup>	5.31 ± 0.39 <sup>a</sup>
K	6.22 ± 0.36 <sup>a</sup>	6.19 ± 0.04 <sup>a</sup>	6.33 ± 0.12 <sup>a</sup>	6.34 ± 0.31 <sup>a</sup>
Total herb (2-cycles)				
N	253.63 ± 5.71 <sup>a</sup>	257.61 ± 12.76 <sup>a</sup>	259.07 ± 10.05 <sup>a</sup>	257.84 ± 9.01 <sup>a</sup>
P	9.57 ± 0.22 <sup>a</sup>	9.76 ± 0.40 <sup>a</sup>	9.83 ± 0.28 <sup>a</sup>	9.89 ± 0.63 <sup>a</sup>
K	11.86 ± 0.39 <sup>a</sup>	11.98 ± 0.35 <sup>a</sup>	12.14 ± 0.08 <sup>a</sup>	12.18 ± 0.46 <sup>a</sup>
System total				
N	527.33 ± 14.37 <sup>a</sup>	535.40 ± 21.13 <sup>a</sup>	544.83 ± 13.40 <sup>a</sup>	538.57 ± 4.95 <sup>a</sup>
P	57.39 ± 0.99 <sup>a</sup>	61.44 ± 2.51 <sup>a</sup>	62.21 ± 3.30 <sup>a</sup>	62.88 ± 3.70 <sup>a</sup>
K	34.87 ± 1.84 <sup>a</sup>	34.55 ± 1.0 <sup>a</sup>	35.38 ± 0.91 <sup>a</sup>	35.23 ± 1.21 <sup>a</sup>

Data are represented as mean ± SE.

<sup>a,s</sup> Values within the same row with different superscript letters are significantly different ( $p < 0.05$ ).

N, nitrogen; P, phosphate; K, potassium.

noticeable drop in water temperature at weeks 4 and 6, and this could be attributed to substantial water replenishment in the culture tanks due to water loss. Water loss in aquaponic systems could be a result of leaf transpiration and evaporation from the system (Liang & Chien, 2013).

Most importantly, the ammonia level in this study suggests that the organisms raised in the polyculture aquaponics system may have had no risk of ammonia toxicity. Consequently, this explains the similarity in the survival of the experimental fishes as observed in the various treatments. In a similar case, the survival and growth of hybrid lemon fin barb and red claw crayfish in this

study were not significantly affected by the different durations of lighting provided for the *O. basilicum*. This observation is in line with the findings of El-Sayed & Kawanna (2004), who observed that there were no differences in the growth performance of Nile tilapia fingerlings reared under four different photoperiods, and Ogah et al. (2020b), who observed no differences in the growth of hybrid lemon fin barb reared with peppermint exposed to different durations of night lighting in a polyculture aquaponic system. It should be noted that the fish in this study and those of Ogah et al. (2020b) were not directly exposed to the different prolonged lighting. Hence, this could be the reason

**Table 7. Nutrient recovery (%) of hybrid lemon fin barb, red claw crayfish, and *Ocimum basilicum* exposed to different night light regimen in a polyculture aquaponic system**

% Recovery	0 h	4 h	8 h	12 h
Fish				
N	30.94 ± 1.83 <sup>a</sup>	31.20 ± 0.67 <sup>a</sup>	31.61 ± 1.90 <sup>a</sup>	30.81 ± 0.58 <sup>a</sup>
P	44.58 ± 2.38 <sup>a</sup>	48.16 ± 0.27 <sup>a</sup>	48.30 ± 2.11 <sup>a</sup>	48.27 ± 1.78 <sup>a</sup>
K	37.63 ± 1.43 <sup>a</sup>	37.04 ± 0.84 <sup>a</sup>	37.73 ± 0.17 <sup>a</sup>	37.06 ± 1.55 <sup>a</sup>
Crayfish				
N	18.69 ± 0.61 <sup>a</sup>	19.40 ± 0.81 <sup>a</sup>	20.14 ± 1.71 <sup>a</sup>	20.48 ± 0.96 <sup>a</sup>
P	9.32 ± 0.39 <sup>a</sup>	9.70 ± 0.35 <sup>a</sup>	9.70 ± 0.50 <sup>a</sup>	10.25 ± 0.67 <sup>a</sup>
K	6.07 ± 0.18 <sup>a</sup>	6.23 ± 0.63 <sup>a</sup>	6.36 ± 0.81 <sup>a</sup>	6.57 ± 0.81 <sup>a</sup>
Total (fish and crayfish)				
N	49.64 ± 1.95 <sup>a</sup>	50.60 ± 0.45 <sup>a</sup>	51.75 ± 1.26 <sup>a</sup>	51.28 ± 0.39 <sup>a</sup>
P	53.89 ± 1.99 <sup>a</sup>	57.87 ± 0.53 <sup>a</sup>	58.01 ± 1.62 <sup>a</sup>	58.51 ± 2.20 <sup>a</sup>
K	43.70 ± 1.59	43.27 ± 1.41 <sup>a</sup>	44.10 ± 0.66 <sup>a</sup>	43.63 ± 0.86 <sup>a</sup>
<i>O. basilicum</i> (cycle 1)				
N	11.39 ± 0.78 <sup>a</sup>	11.75 ± 0.39 <sup>a</sup>	11.64 ± 0.32 <sup>a</sup>	11.26 ± 0.53 <sup>a</sup>
P	3.78 ± 0.28 <sup>a</sup>	4.11 ± 0.10 <sup>a</sup>	4.05 ± 0.08 <sup>a</sup>	3.98 ± 0.32 <sup>a</sup>
K	7.71 ± 0.44 <sup>a</sup>	7.85 ± 0.27 <sup>a</sup>	7.76 ± 0.23 <sup>a</sup>	7.63 ± 0.61 <sup>a</sup>
<i>O. basilicum</i> (cycle 2)				
N	12.86 ± 0.98 <sup>a</sup>	12.95 ± 0.98 <sup>a</sup>	12.73 ± 0.41 <sup>a</sup>	12.41 ± 1.00 <sup>a</sup>
P	4.51 ± 0.43 <sup>a</sup>	4.70 ± 0.36 <sup>a</sup>	4.61 ± 0.13 <sup>a</sup>	4.63 ± 0.50 <sup>a</sup>
K	8.51 ± 0.66 <sup>a</sup>	8.43 ± 0.43 <sup>a</sup>	8.45 ± 0.06 <sup>a</sup>	8.30 ± 0.72 <sup>a</sup>
Total herb (2-cycles)				
N	24.25 ± 1.74 <sup>a</sup>	24.70 ± 1.29 <sup>a</sup>	24.37 ± 0.73 <sup>a</sup>	23.66 ± 1.52 <sup>a</sup>
P	8.29 ± 0.71 <sup>a</sup>	8.81 ± 0.46 <sup>a</sup>	8.66 ± 0.19 <sup>a</sup>	9.00 ± 0.58 <sup>a</sup>
K	16.23 ± 0.92 <sup>a</sup>	16.29 ± 0.61 <sup>a</sup>	16.21 ± 0.17 <sup>a</sup>	15.92 ± 1.28 <sup>a</sup>
System total				
N	73.89 ± 3.54 <sup>a</sup>	75.30 ± 1.49 <sup>a</sup>	76.13 ± 0.89 <sup>a</sup>	74.95 ± 1.13 <sup>a</sup>
P	62.19 ± 2.68 <sup>b</sup>	66.68 ± 0.90 <sup>a</sup>	66.67 ± 1.68 <sup>b</sup>	67.12 ± 2.66 <sup>b</sup>
K	59.92 ± 2.19 <sup>a</sup>	59.56 ± 1.96 <sup>a</sup>	60.30 ± 0.79 <sup>a</sup>	59.55 ± 2.11 <sup>a</sup>

Data are represented as mean ± SE.

<sup>ab</sup>Values within the same row with different superscript letters are significantly different ( $p < 0.05$ ).

N, nitrogen; P, phosphate; K, potassium.

for the disparity in the performance evaluation compared with those of Liang & Chien (2015) who reported improved growth performance of tilapia and loach exposed directly to prolonged night lighting in a raft aquaponic system.

According to Avgoustaki (2019), one of the most important factors for evaluating growth conditions and consumer choices for vegetables and herbs is plant quality. The indices of plant quality as opined by researchers include colour, shape, height, size (morphometry) of herb leaves, and yield (McElroy et al., 2004; Miguel et al., 2002; Zhang et al., 2018). In conventional farming, photoperiod plays an important role in the size/area of

leaves and biomass (Adams & Langton, 2005). The assumption at the beginning of the current study was that an increase in the supplementary night lighting (or illumination period) would result in a significant increase in yield compared to the *O. basilicum* grown under control. However, the result of the current study is at variance with this assumption as it is observed that there is no significant difference in all morphometry parameters taken from the *O. basilicum* grown under the increased illumination periods. Similarly, McElroy et al. (2004) had earlier reported that the root weights of Florida betony exposed to long-day and short-day photoperiods were similar after 10 weeks of

culture. In contrast, the study by Ogah et al. (2020b) observed a 19% higher yield, stem length, and leaf numbers in peppermint plants exposed to 12 h additional night lighting in a polyculture aquaponic system when compared with the control group. Also, Miguel et al. (2002) reported higher fresh mass yields of several medicinal herbs when they were exposed to a longer day length. The observed differences in the responses of the different plants to additional night lighting may be due to the differences in their biology and photoperiodic preferences. Nonetheless, Zhang et al. (2018) had earlier hypothesized that the differences in the ability of different plants to use larger amounts of carbohydrates for physiological metabolism activities/growth could explain the diversity of performance of plants under different photoperiods.

The trends of NPK recoveries by red claw crayfish, hybrid lemon fin barb, and the *O. basilicum* in this experiment were similar irrespective of the different supplementary night lighting regimes to which the *O. basilicum* were subjected. Whiteman & Room (1991) had earlier stated that temperature may have a direct effect on the nutrient absorption capacity of organisms. However, the additional lighting was not observed to increase the temperature of the system or environment, which may partly justify the similarities in the NPK recoveries of the treatments and control groups. However, Ogah et al. (2020b) had earlier reported that additional night lighting did not affect growth and NPK recoveries of hybrid lemon fin barb, and their finding is in contrast with the present study, which showed an 18% nutrient uptake of the peppermint plant exposed to the prolonged night lighting. Differences in the biology and photoperiod preferences of the two plants used may explain the performance differences. The recovery of 0.04 % P and 1.68 % K in cucumber co-cultured with common carp in an aquaponic system was reported by Guzel et al. (2018), while Ogah et al. (2020b) reported 4.54%–8.15% N, 0.94%–1.91% P and 2.06%–2.31% K recovery rate by peppermint in a similar aquaponic system. These reports show that *O. basilicum* in the present study has a higher NPK recovery than the reported plants. A study by Moustafa et al. (2020), which reported 25.8% N and 2.74% P recovery in a RAS with Nile tilapia, was lower compared to the observations of this study for fish (30.81%–31.2% N; 44.58%–48.3% P) and crayfish (18.96%–20.48% N; 9.32%–10.25% P). The differences in nutrient recoveries may be linked to the different culture systems used in both studies.

## Conclusion

The findings of this study showed that varying supplementary

night lighting has no significant effect on the growth performance of *O. basilicum*, hybrid lemon fin barb, as well as red claw crayfish. A similar observation was made with respect to their nutrient recoveries and water qualities. Future studies should also investigate the effect of exposing the fish directly to different light regimes (such as light intensity, source, and quality) on the performance of fish and plants co-cultured in the aquaponic system.

## Competing interests

No potential conflict of interest relevant to this article was reported.

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## Availability of data and materials

Upon reasonable request, the datasets of this study can be available from the corresponding author.

## Ethics approval and consent to participate

The authors confirm that the ethical policies of the journal have been adhered to and the appropriate ethical review committee approval has been received. The US National Research Council's guidelines for the Care and Use of Laboratory Animals were followed.

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## References

- Adams SR, Langton FA. Photoperiod and plant growth: a review. *J Hortic Sci Biotechnol*. 2005;80:2-10.
- Ahmed AF, Attia FAK, Liu Z, Li C, Wei J, Kang W. Antioxidant activity and total phenolic content of essential oils and extracts of sweet basil (*Ocimum basilicum* L.) plants. *Food Sci Hum Wellness*. 2019;8:299-305.
- Association of Official Analytical Chemists [AOAC]. Changes in Official Methods of Analysis Made at the Annual Meeting. In: Cunniff PA, editor. *Official Methods of Analysis of AOAC International*. 16 ed. Arlington, TX: AOAC; 1997.
- Avgoustaki DD. Optimization of photoperiod and quality assessment of basil plants grown in a small-scale indoor cultivation system for reduction of energy demand. *Energies*. 2019;12:3980.
- Bader N. Sample preparation for flame atomic absorption spectroscopy: an overview. *Rasāyan J Chem*. 2011;4:49-55.
- Boyd CE, Gross A. Water use and conservation for inland aquaculture ponds. *Fish Manag Ecol*. 2000;7:55-63.
- Dou H, Niu G, Gu M, Masabni JG. Responses of sweet basil to different daily light integrals in photosynthesis, morphology, yield, and nutritional quality. *HortScience*. 2018;53:496-503.
- Eck M, Körner O, Jijakli MH. Nutrient cycling in aquaponics systems. In: Goddek S, Joyce A, Kotzen B, Burnell GM, editors. *Aquaponics food production systems: combined aquaculture and hydroponic production technologies for the future*. Cham: Springer; 2019. p. 231-46.
- El-Sayed AFM, Kawanna M. Effects of photoperiod on the performance of farmed Nile tilapia *Oreochromis niloticus*: I. growth, feed utilization efficiency and survival of fry and fingerlings. *Aquaculture*. 2004;231:393-402.
- Fernández-Cabanás VM, Pérez-Urrestarazu L, Juárez A, Kaufman NT, Gross JA. Comparative analysis of horizontal and vertical decoupled aquaponic systems for basil production and effect of light supplementation by LED. *Agronomy*. 2020;10:1414.
- Guzel S, Odun UC, Cakmakclı T, Cakmakclı O, Sahin U. The effect of cucumber (*Cucumis sativus*) cultivation in aquaponic and hydroponic systems on plant nutrient elements and antioxidant enzyme activity. *Fresenius Environ Bull*. 2018;27:553-8.
- Hasan Z, Dhahiyat Y, Andriani Y, Zidni I. Water quality improvement of Nile tilapia and catfish polyculture in aquaponics system. *Nus Biosci*. 2017;9:83-5.
- Liang JY, Chien YH. Effects of feeding frequency and photoperiod on water quality and crop production in a tilapia-water spinach raft aquaponics system. *Int Biodeterior Biodegrad*. 2013;85:693-700.
- Liang JY, Chien YH. Effects of photosynthetic photon flux density and photoperiod on water quality and crop production in a loach (*Misgurnus anguillicandatus*)–nest fern (*Asplenium nidus*) raft aquaponics system. *Int Biodeterior Biodegrad*. 2015;102:214-22.
- Love DC, Fry JP, Li X, Hill ES, Genello L, Semmens K, et al. Commercial aquaponics production and profitability: findings from an international survey. *Aquaculture*. 2015;435:67-74.
- Malayeri SH, Hikosaka S, Ishigami Y, Goto E. Growth and photosynthetic rate of Japanese mint (*Mentha arvensis*) grown under controlled environment. *VI Intern Symp Light Hortic*. 2011;907:73-9.
- Martan E. Polyculture of fishes in aquaponics and recirculating aquaculture. *Aquaponics J*. 2008;48:28-33.
- McElroy JS, Yelverton FH, Neal JC, Ruffy TW Jr. Influence of photoperiod and temperature on vegetative growth and development of Florida betony (*Stachys floridana*). *Weed Sci*. 2004;52:267-70.
- Miguel MG, Duarte F, Venâncio F, Tavares R, Guerrero C, Martins H, et al. Changes of the chemical composition of the essential oil of portuguese *Thymus mastichina* in the course of two vegetation cycles. *Acta Hortic*. 2002:83-6.
- Moustafa YTA, Ali A, Gomha SAA. Nitrogen and phosphorus budget for Nile tilapia hatchery. *Egypt J Aquac*. 2020;10:1-22.
- Nuwansi KKT, Verma AK, Tiwari VK, Prakash C, Chandrakant MH. Standardization of the stocking density ratios of koi carp (*Cyprinus carpio* var. koi): goldfish (*Carassius auratus*) in polyculture aquaponic recirculating system. *Turk J Fish Aquat Sci*. 2017;17:1271-8.
- Ogah SI, Kamarudin MS, Nurul-Amin SM, Puteri Edaroyati MW. Nutrient recycling through aquaponics and night-lighting. *J Environ Biol*. 2020b;41:1113-25.
- Ogah SI, Kamarudin MS, Nurul Amin SM, Wahab Puteri Edaroyati M. Biological filtration properties of selected herbs in an aquaponic system. *Aquac Res*. 2020a;51:1771-9.
- Oladimeji AS, Olufeagba SO, Ayuba VO, Sololmon SG, Okomodua VT. Effects of different growth media on water quality and plant yield in a catfish-pumpkin aquaponics system.

- J King Saud Univ Sci. 2020;32:60-6.
- Oommen AK, Saji A, Joseph S, Kuriakose BP. Automated water quality monitoring system for aquaponics. *Int Res J Eng Technol*. 2019;6:7832-41.
- Ouzounis T, Rosenqvist E, Ottosen CO. Spectral effects of artificial light on plant physiology and secondary metabolism: a review. *HortScience*. 2015;50:1128-35.
- Petrucio MM, Esteves FA. Influence of photoperiod on the uptake of nitrogen and phosphorus in the water by *Eichhornia crassipes* and *Salvinia auriculata*. *Rev Bras Biol*. 2000;60:373-9.
- Rakocy JE. Aquaponics—integrating fish and plant culture. *Aquac Prod Syst*. 2012:344-86.
- Rakocy JE, Masser MP, Losordo TM. Recirculating aquaculture tank production systems: aquaponics—integrating fish and plant culture [Internet]. SRAC. 2006 [cited 2023 Jun 5]. <https://fisheries.tamu.edu/files/2013/09/SRAC-Publication-No.-454-Recirculating-Aquaculture-Tank-Production-Systems-Aquaponics-Integrating-Fish-and-Plant-Culture.pdf>
- Whiteman JB, Room PM. Temperatures lethal to *Salvinia molesta* Mitchell. *Aquat Bot*. 1991;40:27-35.
- Yildiz HY, Robaina L, Pirhonen J, Mente E, Domínguez D, Parisi G. Fish welfare in aquaponic systems: its relation to water quality with an emphasis on feed and faeces—a review. *Water*. 2017;9:13.
- Zhang X, He D, Niu G, Yan Z, Song J. Effects of environment lighting on the growth, photosynthesis, and quality of hydroponic lettuce in a plant factory. *Int J Agric Biol Eng*. 2018;11:33-40.